

α -MSH and neurofilament M-protein share a continuous epitope but not extended sequences

An explanation for neurofibrillary staining with α -MSH antibodies

Gerry Shaw, Stefan Fischer and Klaus Weber

Max Planck Institute for Biophysical Chemistry, D-3400 Göttingen, FRG

Received 2 January 1985

The long recognized neurofibrillary immunoreactivity of the nervous system with α -MSH antibodies arises from an epitope on neurofilament M-protein which we have now characterized. It is situated in the amino terminal residues where M-protein and α -MSH exhibit similar but not identical sequences. Their divergence past residue 5 precludes a physiological significance of the crossreactivity which seems to have arisen fortuitously. Our results question previous speculations as to the existence of extrapituitary α -MSH-like hormones.

α -MSH Neuropeptide Neurofilament Neuron Epitope Crossreactivity

1. INTRODUCTION

The main source of α -melanocyte-stimulating hormone (α -MSH), an acetyl-tridecapeptide amide, is the pituitary gland where it is processed from a large polypeptide precursor [1]. However, many studies dating back at least to 1977 documented strong neurofibrillary α -MSH immunoreactivity in most parts of the nervous system (see, e.g., [2–6]). Radioimmunoassays were used to quantitate this material and various speculations as to extrapituitary α -MSH-related hormones have been advanced. The neurofibrillary specificity of the reaction was partially explained when Dräger et al. [2] found that α -MSH antibodies stain neurofilaments, the neuronal form of intermediate filaments. Immunoblotting revealed reactivity only for the middle-sized M-protein, one of the three filament components: L (68 kDa), M (160 kDa), and H (200 kDa) [7]. This and previous observations [2–6] raised the possibility that neurofilaments may contain peptide hormone-like sequences, and we note that intermediate filament proteins have been postulated to be related to

steroid hormone receptors [8]. With the recent availability of partial neurofilament protein sequences [9,10], we felt in a good position to follow up these earlier findings.

2. MATERIALS AND METHODS

Chicken desmin, porcine neurofilament M-protein, and fragments of M covering residues 1–102 (headpiece) and the carboxy-terminal tailpiece extension were isolated [9,10]. Rabbit antiserum to α -MSH was obtained from Immunonuclear Corp. (Stillwater, MN, USA). Initial studies used lot no.31261 as in [2]. As this serum was later not available we made use of lot no. 8410018. Competitive ELISA between immobilized M-protein, α -MSH antibody and various peptides was performed as described for another antigen-antibody system [11]. Western blotting was performed as in [12]. Peroxidase-conjugated second antibodies were used. α -MSH, desacetyl α -MSH, ACTH, ACTH_{1–10}, and the tetrapeptide S-Y-S-M were from Bachem (Bubendorf, Switzerland). Serine-tyrosine was obtained from

Serva. Acetylation was performed with acetic anhydride using standard techniques. Acetylated peptides were purified by HPLC and characterized by amino acid composition and electrophoresis. Tobacco mosaic virus (TMV) coat protein and a rabbit antiserum known to react with the amino-terminal 10 residues of this protein [13] were kindly donated by Dr Van Regenmortel, CNRS, Strasbourg, France. Sequences are given using the one-letter system. Ac refers to the *N*-acetyl group.

3. RESULTS AND DISCUSSION

Fig.1 confirms the specific recognition of porcine neurofilament M-protein by anti- α -MSH serum reported by Dräger et al. [2]. Current sequence data on porcine M-protein do not cover the large carboxy-terminal CNBr fragment (tailpiece) although the amino-terminal 436 residues are known [9]. Western blots of cyanogen bromide-cleaved M-protein showed no reactivity of the large carboxy-terminal fragment or the fragment corresponding to residues 68–285 (fig.1). Strong decoration of an 8-kDa fragment indicated the

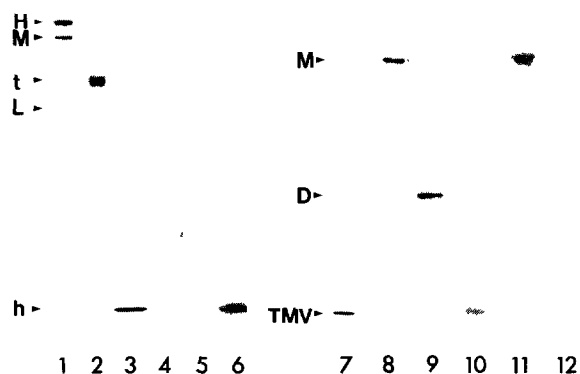


Fig.1. Lanes 1–6, 12.5% polyacrylamide gel (1–3) and corresponding Western blot (4–6) of purified neurofilament proteins (H, M and L), M-protein tail piece (t) and M-protein head piece (h) with α -MSH antibody. Both the intact protein (lanes 1 and 4) and the small amino-terminal moiety (lanes 3 and 6) are strongly stained. Lanes 7–12, 10% polyacrylamide gel (7–9) and corresponding Western blot (10–12) of TMV coat protein (TMV), neurofilament M-protein (M), and chicken desmin (D). Blot was stained with α -MSH antibody. α -MSH antibody recognizes M and TMV, but not chicken desmin.

amino-terminal fragment (residues 1–67) which was corroborated using a pure preparation of a proteolytic fragment covering residues 1–102 (headpiece). Having localized the α -MSH-like antigen to residues 1–67, a sequence comparison with α -MSH indicated only one candidate for a continuous epitope, i.e., the amino-terminal residues. These are Ac-S-Y-T-L-D- and Ac-S-Y-S-M-E- in M-protein and α -MSH, respectively (see table 1).

A modified ELISA with carrier-bound porcine M-protein was used to test various peptides for antigen competition. α -MSH consists of the first 13 residues of ACTH but is acetylated at the amino terminus and amidated at the carboxyl end. The much lower reactivity of ACTH and desacetylated α -MSH in comparison to α -MSH implicated the acetylated end. The amino-terminal tetrapeptide S-Y-S-M displayed very poor competition under our assay conditions, whereas its *N*-acetyl derivative showed significant reaction (table 1). The *N*-acetylated dipeptide S-Y exhibited slight inhibition at very high concentrations, though S-Y alone showed no inhibition. Acetylated ACTH_{1–10} inhibited about as strongly as α -MSH, whereas the non-acetylated form inhibited much more weakly. Both lots of α -MSH antibody we examined behaved in an almost identical manner in these experiments. A previous study made use of four different α -MSH antisera, all of which exhibited staining of non-pituitary structures [6], suggesting that many α -MSH antibodies may be able to recognize the M-protein.

Purified chicken desmin (fig.1, table 1) is not recognized, indicating a requirement for tyrosine at position 2. The combined results suggest that strong antibody binding requires acetyl-serine-tyrosine, followed by at least two and preferably more amino acids the identity of which can, at least within limits, be somewhat variable. The identification of the antigenic site on the M-protein explains the neuronal specificity of α -MSH staining seen in histological samples, since various intermediate filament [9] and other cytoskeletal proteins (see below) differ from M in their amino-terminal sequence [9].

We searched the National Biomedical Research Foundation protein sequence data bank for other proteins exhibiting an acetylated amino-terminal serine followed by a tyrosine and a serine or threonine. This requirement was fulfilled by TMV

Table 1

Summary of results from competitive ELISA and Western blotting experiments

Peptide used in competitive ELISA	Concentration required for 50% inhibition
S-Y	> 333 μM ^a
Ac-S-Y	300 μM
S-Y-S-M (ACTH ₁₋₄)	> 100 μM ^a
Ac-S-Y-S-M (acetylated ACTH ₁₋₄)	235 nM
S-Y-S-M-E-H-F-R-W-G-K-P-V-amide (desacetyl α -MSH)	60 μM
Ac-S-Y-S-M-E-H-F-R-W-G-K-P-V-amide (α -MSH)	380 pM

N-terminus of protein used in Western blotting experiments	Result of Western blotting experiment
Ac-S-Y-T-L-D-S-L-G-N-P-S-S-A... (M protein)	++
Ac-S-Y-S-I-T-T-P-S-Q-F-V-F-L... (TMV coat protein)	++
Ac-S-Q-S-Y-S-S-S-Q-R-V-S-S-Y... (chicken desmin)	—

^a The peptide failed to reach 50% inhibition at the highest concentration employed

coat protein. The sequence Ac-S-Y-S-I-T- [14,15] shows good homology to α -MSH and M-protein for the first four residues. Western blotting (fig.1) revealed that TMV coat protein was also reactive with α -MSH antibodies. In addition, a rabbit antiserum known to recognize the amino-terminal 10 residues of coat protein [13] recognizes M-protein on Western blots, although very weakly (not shown).

The amino-terminal region of TMV coat protein belongs to the increasing list of linear or continuous epitopes thought to be prime antigenic targets because of their segmental mobility as revealed by X-ray crystallography [13,16]. Such sites are very often situated in loops and can also occur at the termini of different proteins [11,13,16]. Current concepts of intermediate filament structure based on biochemical and sequence considerations already suggest the presence of loops in the amino-terminal region of M-protein as in other intermediate filament proteins [9,10]. The crossreactivity of α -MSH antibodies documented here confirms the general principle of a linear epitope situated in a flexible loop at the amino-terminal end. Tolerable sequence variations in this region account for the crossreactivity of the α -MSH antibodies with proteins as divergent as α -MSH, neurofilament M-protein, and TMV coat protein. As the sequence relationship between α -

MSH and neurofilament M-protein is lost past residue 5 (see table 1) we conclude that neurofilament proteins cannot serve as a precursor or storage form of α -MSH-like hormones. This should lay to rest previous speculations on extrapituitary α -MSH-like hormones in the nervous system [3–6] based solely on immunological crossreactivity.

REFERENCES

- [1] Douglass, J., Civilli, O. and Herbert, E. (1984) *Annu. Rev. Biochem.* 53, 665–725.
- [2] Dräger, U.C., Edwards, D.L. and Kleinschmidt, J. (1983) *Proc. Natl. Acad. Sci. USA* 80, 6408–6412.
- [3] Swaab, D.F. and Fisser, B. (1977) *Neurosci. Lett.* 7, 313–317.
- [4] Vaudry, H., Tonon, M.C., Delarue, R., Vailant, R. and Kraicer, J. (1978) *Neuroendocrinology* 27, 9–24.
- [5] O'Donahue, T.L., Miller, R.L. and Jacobowitz, D.M. (1979) *Brain Res.* 176, 101–123.
- [6] Watson, S.J. and Akil, H. (1980) *Brain Res.* 182, 217–223.
- [7] Hoffmann, P.N. and Lasek, R.J. (1975) *J. Cell Biol.* 66, 351–366.
- [8] Traub, P., Nelson, W.J., Kuehn, S. and Vorgias, C.E. (1983) *J. Biol. Chem.* 258, 1456–1466.
- [9] Geisler, N., Fischer, S., Vandekerckhove, J., Plessmann, U. and Weber, K. (1984) *EMBO J.* 3, 2701–2706.

- [10] Geisler, N. and Weber, K. (1982) *EMBO J.* 1, 1649–1656.
- [11] Wehland, J., Schroeder, H.C. and Weber, K. (1984) *EMBO J.* 3, 1295–1300.
- [12] Shaw, G., Debus, E. and Weber, K. (1984) *Eur. J. Cell Biol.* 34, 130–136.
- [13] Westhof, E., Altschuh, D., Moras, D., Bloomer, A.C., Mondragon, A., Klug, A. and Van Regenmortel, M.H.V. (1984) *Nature* 311, 123–126.
- [14] Anderer, F.A., Uhlig, H., Weber, E. and Schramm, B. (1960) *Nature* 186, 922–925.
- [15] Tsugita, A., Gish, D.T., Young, J., Fraenkel-Conrat, H., Knight, C.A. and Stanley, W.M. (1960) *Proc. Natl. Acad. Sci. USA* 46, 1463–1469.
- [16] Tainer, J.A., Getzoff, E.D., Alexander, H., Houghton, R.A., Olson, A.J., Lerner, R.A. and Hendrickson, W.A. (1984) *Nature* 312, 127–134.